

Review

# Cross-talk of opioid peptide receptor and $\beta$ -adrenergic receptor signalling in the heart

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Received 8 February 2004; received in revised form 20 April 2004; accepted 21 April 2004

Time for primary review 21 days

## Abstract

Opioid peptide receptor (OPR) and  $\beta$ -adrenergic receptor ( $\beta$ -AR) are well-established members of G-protein-coupled receptor (GPCR) superfamily and are involved in regulating cardiac contractility, energy metabolism, myocyte survival or death. OPRs are typical  $G_i/G_o$ -coupled receptors and activated by opioid peptides derived from the endorphin, dynorphin and enkephalin families, whereas  $\beta$ -AR stimulated by catecholamines is the model system for  $G_s$ -coupled receptors. While it is widely accepted that  $\beta$ -AR stimulation serves as the most powerful means to increase cardiac output in response to stress or exercise, we have only begun to appreciate functional roles of OPR stimulation in regulating cardiovascular performance. Cardiovascular regulatory effects of endogenous opioids were initially considered to originate from the central nervous system and involved the pre-synaptic co-release of norepinephrine with enkephalin from sympathetic neuronal terminals in the heart. However, opioid peptides of myocardial origin have been shown to play important roles in local regulation of the heart. Notably, OPR stimulation not only inhibits cardiac excitation–contraction coupling, but also protects the heart against hypoxic and ischemic injury via activation of  $G_i$ -mediated signalling pathways. Further, OPRs functionally and physically cross-talk with  $\beta$ -ARs via multiple hierarchical mechanisms, including heterodimerization of these receptors, counterbalance of functional opposing G protein signalling, and interface at downstream signalling events. As a result, the  $\beta$ -AR-mediated positive inotropic effect and increase in cAMP are markedly attenuated by OPR activation in isolated cardiomyocytes as well as sympathectomized intact rat hearts. This brief review will focus on the interaction between  $\beta$ -AR and OPR and its potential physiological and pathophysiological relevance in the heart.

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**Keywords:** G protein-coupled receptors;  $\beta$ -Adrenergic receptors; Opioid peptide receptors; Receptor dimerization; Cardiac contractility; Cardiac preconditioning

## 1. Introduction

The Human Genome Project has demonstrated that the family of G-protein-coupled receptors (GPCRs) is the largest and most diverse gene family in the human genome [1]. The GPCR superfamily is involved in the transduction of stimulatory or inhibitory signals in response to a wide

array of stimuli, and has also long been considered as a most important therapeutic target. Increasing evidence has shown that members of GPCR superfamily that couple to different classes of G proteins are co-expressed in numerous tissues and interact with each other at multiple levels, including receptor, G protein, or downstream signalling pathways, thus altering signalling and trafficking properties of these receptors. In the heart,  $\beta$ -adrenergic receptors ( $\beta$ -ARs) and opioid peptide receptors (OPRs) are co-expressed, and are coupled to functionally opposite G protein families,  $G_s$  and  $G_{i/o}$ , respectively. Emerging evidence suggests that  $\beta_2$ -AR and OPRs can activate more than one G protein family, as manifested by dual coupling of  $\beta_2$ -AR to  $G_s$  and

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$G_i$  and OPR- $\delta$  to  $G_i$  and  $G_q$ . The “cross-talk” between OPRs with  $\beta$ -ARs not only antagonizes  $\beta$ -AR-mediated positive inotropic effect, but also alters these receptors’ trafficking and signalling characteristics. In this brief review, we intend to highlight the key features of opioid peptides and OPRs in the heart and their interactions with  $\beta$ -AR signalling.

## 2. Endogenous opioid peptides: from brain to heart

The pentapeptides methionine-enkephalin and leucine-enkephalin were first discovered in the brain and adrenal gland [2,3]. Shortly after, enkephalin-containing peptides and OPRs were identified throughout the central and peripheral nervous system, including the afferent neurons terminating in the heart [4,5]. Three gene products, pro-opiomelanocortin, prodynorphin, and proenkephalin, are the precursors for endorphins, dynorphins, and enkephalins, respectively [6,7]. The neural signalling of opioid peptides has been well characterized in multiple roles, including bradycardia, tachycardia, hypertension and hypotension [8–12]; however, this has overshadowed the functional roles of endogenous opioids of cardiac origin. Over the past decade, accumulating evidence demonstrates that both pro-enkephalin and prodynorphin and their final products are expressed and produced directly by cardiomyocytes from mammalian species, including rat, guinea pig, and dog (for review, see Ref. [13]), although opioid peptides are also produced and released in the central nervous system and from peripheral neuronal terminals in the heart where they are co-released with catecholamines [14].

Specifically, a large proportion of enkephalins in the heart appear to be produced by cardiomyocytes [15]. Expression of the proenkephalin gene results in a 31-kDa polypeptide precursor that after post-translational processing yields methionine- and leucine-enkephalin, the heptapeptide methionine-enkephalin-Arg6-Phe7 (MEAP), the octapeptide methionine-enkephalin-Arg6-Gly7-Leu8, and several larger peptides, such as peptides B, E, F, I, and BAM 20P [7]. Proenkephalin mRNA is highly expressed in adult rat heart, particularly in the left ventricle [16]. Although the expression of cardiac proenkephalin mRNA is higher than that in brain [17], the abundance of extracted proenkephalin-derived peptides are much lower in the heart [18]. Interestingly, 95% of enkephalins recovered from rat cardiac ventricle are concentrated in precursor and large intermediate peptides such as proenkephalin (20%), Peptide B (43%), and MEAP (32%) rather than in the smaller end products [19]. Whilst the function of these larger precursor proteins is unclear, they might serve as a ready reservoir or precursor for cleavage into the smaller OPR-active peptides, thereby resulting in augmented release under certain pathophysiological conditions. For instance, leucine-enkephalin release into the coronary sinus effluent in rats is significantly increased in response to ischemia–reperfusion injury [20].

In addition to enkephalins, cardiomyocytes also synthesize and secrete dynorphin B, a biologically active end product of the prodynorphin gene [21], which binds specifically to  $\kappa$ -OPR [22] (see below). It is noteworthy that there is an intrinsic *autocrine loop* regulation of the prodynorphin gene expression. Specifically, dynorphin B, the end product of the prodynorphin gene, stimulates  $\kappa$ -OPR and serves as a positive feedback to augment the prodynorphin gene expression via a mechanism involving translocation of PKC- $\alpha$  into nuclei and subsequent activation of nuclear PKC- $\delta$  and - $\epsilon$  [23]. Multiple lines of evidence reveal that prodynorphin gene expression and dynorphin B abundance are markedly increased in cardiomyopathic hearts from Syrian hamsters of the BIO14.6 strain, perhaps largely due to cardiomyopathy-associated abnormalities in  $Ca^{2+}$  handling and enhanced activation of PKC [24,25]. Together, these studies strongly support the perception that the heart is a complex endocrine organ, and that myocardial function might be particularly affected by opioid peptides in an autocrine or paracrine manner.

## 3. Opioid peptide receptors and modulation of cardiac excitation–contraction coupling

Three genetically and pharmacologically distinct OPR subtypes,  $\mu$ ,  $\kappa$ , and  $\delta$ , have been identified [26–30]. Although these closely related OPR subtypes share about 50% amino acid sequence homology and some functional similarities [31], they differ in their ligand binding and tissue distribution. Specifically,  $\delta$ -OPRs have the highest affinity for enkephalins;  $\kappa$ -OPRs bind preferentially to dynorphins; and  $\mu$ -OPRs are selectively sensitive to endorphins, including morphine [32]. Further, it has been demonstrated that  $\delta$ - and  $\kappa$ -OPRs, but not  $\mu$ -OPRs, are present in adult rat ventricular myocardium [9,33–38]. In contrast, only  $\mu$ - and  $\kappa$ -OPRs are observed in neonatal hearts [37]. Thus, there might be a development-dependent expression of  $\delta$ -OPR in the heart. Under certain pathological circumstances, OPR subtype expression can be up- or down-regulated. Moreover, each OPR subtype can be further subdivided. While  $\delta_1$ - and  $\delta_2$ -OPR subtypes have been pharmacologically identified [37,39], only one  $\delta$ -OPR has so far been cloned [26]. Thus, the existence and significance of  $\delta_1$ - and  $\delta_2$ -OPR subtypes merits further investigation. In addition, new subtypes for  $\kappa$ - and  $\mu$ -OPR have also been reported, but they are yet to be identified in heart.

Importantly, activation of  $\delta$ -OPRs directly modulate systemic vascular resistance in intact organisms. In adult rat ventricular myocytes, OPR stimulation suppresses the L-type  $Ca^{2+}$  current [40] and affects sarcoplasmic reticulum (SR)  $Ca^{2+}$  depletion [41], resulting in reduced  $[Ca^{2+}]_i$  transient and contractility [41]. These effects are reversed by the OPR antagonist, naloxone. Stimulation of the closely related OPR subtype,  $\kappa$ -OPR, also exhibits a robust inhibitory effect on cardiac excitation–contraction coupling. In

adult rat ventricular myocytes, the  $\kappa$ -OPR agonist, dynorphin B, causes a transient positive inotropic effect via augmenting  $\text{Ca}^{2+}$  release from SR and intracellular alkalization, and in the steady-state causes a reduction in myocyte contractility due to SR  $\text{Ca}^{2+}$  depletion [41–45]. The inhibitory effects of both  $\delta$ -OPR and  $\kappa$ -OPR on cardiac excitation–contraction coupling are likely mediated by multiple G protein signalling pathways such as  $G_{i/o}$  and  $G_q$ , as evidenced by their sensitivity to pertussis toxin (PTX, a  $G_{i/o}$  inhibitor) and activation of phospholipase C/PKC pathway [41–45].

#### 4. Heterodimerization of $\beta$ -ARs with OPRs

Increasing evidence has shown that GPCRs can form homodimers or heterodimers. Recent studies suggest that OPRs are capable of forming heterodimers with not only other OPR-subtypes [46,47] but also with  $\beta$ -ARs [48]. Fully functional OPR subtypes can form heterodimers with each other, resulting in a unique population of receptors in terms of ligand binding and intracellular signalling. Heterodimerization of OPR subtypes has been proposed to facilitate selectivity for co-release of the numerous endogenous opioid agonists, thus representing a complex but powerful regulatory mechanism. However, the degree of complexity is even greater than first thought following the finding that oligomerization of OPR with  $\beta_2$ -AR can occur to alter receptor trafficking and signal transduction. Specifically,  $\beta_2$ -AR can physically associate with both  $\delta$ - and  $\kappa$ -OPRs when they are co-expressed in HEK-293 cells [48]. As a result, activation of  $\delta$ -OPR by etorphine induces  $\beta_2$ -AR internalisation and inhibits  $\beta_2$ -AR-mediated mitogen-activated protein kinase (ERK1/2) activation [48]. It is presently unclear whether  $\beta_1$ -AR forms a heterodimer complex with OPRs.

#### 5. Cross-talk between $\beta$ -AR and OPR signalling in regulating cardiac contractility

In the heart, both  $\beta$ -ARs and OPRs are present on cardiac myocyte sarcolemma, and OPR agonists are co-released with the endogenous  $\beta$ -AR agonist, norepinephrine, from nerve terminals [4]. Activation of  $\delta$ -OPRs not only directly modulates cardiac excitation–contraction coupling, as discussed above, but also markedly inhibits  $\beta$ -AR-mediated positive inotropic effects. For example, leucine-enkephalin ( $10^{-8}$  M, a physiologically relevant concentration) overtly attenuates the effect of  $\beta$ -AR stimulation by norepinephrine to increase left ventricular systolic pressure in the isolated perfused rat heart [49], and abolishes  $\beta$ -AR-induced increases in the  $[\text{Ca}^{2+}]_i$  transient and contractility in single rat ventricular myocytes [50]. These anti-adrenergic effects are reversed by the OPR antagonist, naloxone, and are prevented by PTX pretreatment [49,50].

Similarly, activation of  $\kappa$ -OPR with U50,488H also inhibits the effects of  $\beta$ -AR agonist, norepinephrine (NE), to increase  $[\text{Ca}^{2+}]_i$  transient and contractility in single isolated rat ventricular myocytes [51]. Interestingly, in ischemic rat hearts, the inhibitory effect of  $\kappa$ -OPR stimulation on  $\beta$ -AR signalling is markedly enhanced, thus resulting in a cardiac protection as evidenced by reduced arrhythmia [52]. In contrast, in chronic hypoxic rat hearts [53] or spontaneously hypertensive rat cardiac myocytes [54],  $\kappa$ -OPR-mediated inhibition of  $\beta$ -AR positive inotropic effect is lacking or markedly blunted. The reduced inhibition of  $\beta$ -AR signalling might contribute to ultimate cardiomyopathy and heart failure under those pathological circumstances (for review, see Ref. [55]).

It is noteworthy that there is a difference between  $\beta_1$ - and  $\beta_2$ -AR subtypes with respect to their cross-talk with  $G_i/G_o$ -coupled  $\delta$ -OPRs in adult rat myocardium (see Fig. 1). The  $\delta$ -OPR agonist, leucine-enkephalin, markedly inhibits  $\beta_1$ -AR-mediated positive inotropy [49,50]. In contrast, it has no effect on  $\beta_2$ -AR-mediated increase in cardiac contractility [49], indicating that  $\delta$ -OPR signalling selectively interacts with  $\beta_1$ -AR, but not  $\beta_2$ -AR, in regulating myocardial contractility. Similarly, in cultured neonatal rat cardiomyocytes, the  $\beta_1$ -AR-mediated cAMP accumulation and inotropic and lusitropic effects are all blocked by  $G_i$ -coupled  $M_2$ -muscarinic receptor stimulation with carbachol. However, the  $\beta_2$ -AR-induced cAMP accumulation and the inotropic effect are insensitive to  $M_2$  stimulation by carbachol [56]. Although  $\beta_1$ -AR activation of inotropy has been shown to be blocked by adenosine, endothelin, and angiotensin, it has not yet been determined whether  $\beta_2$ -AR-induced contractile response is also sensitive to activation of these respective  $G_i/G_o$ -coupled receptors in the heart.

The exact mechanism underlying the differential interaction of these  $\beta$ -AR subtypes with  $G_i$ -coupled receptors remains elusive. In this regard, it is speculated that the differential interaction between these  $\beta$ -AR subtypes and  $G_i$ -coupled receptors such as  $\delta$ -OPR and  $M_2$ -muscarinic receptors might be, to some extent, attributed to the differential subcellular localization of these  $\beta$ -AR subtypes. In the absence of agonist stimulation,  $\beta_1$ -ARs are enriched in non-caveolae cell surface membranes, whereas  $\beta_2$ -ARs are predominantly distributed in the caveolae membrane fraction in neonatal cardiomyocytes [57]. It has been shown that unstimulated  $M_2$ -muscarinic receptors co-localize with  $\beta_1$ -ARs, but not  $\beta_2$ -ARs, in non-caveolar cell surface membranes in neonatal cardiomyocytes [58]. This may explain, in part, the differential interactions of  $M_2$ -receptor with  $\beta_1$ -ARs versus  $\beta_2$ -ARs. A similar cell architectural arrangement might account for the selective interactions of  $\delta$ -OPRs with  $\beta_1$ -ARs but not  $\beta_2$ -ARs in the heart. Alternatively, a large body of evidence has indicated that  $\beta_2$ -ARs couple to  $G_i$  proteins in addition to the classic  $G_s$  pathway [59–64]. The additional  $G_i$  coupling of  $\beta_2$ -AR might preclude its interaction with other  $G_i$ -coupled receptors such as  $\delta$ -OPR in regulating cardiac contractility. Altogether, multiple hier-

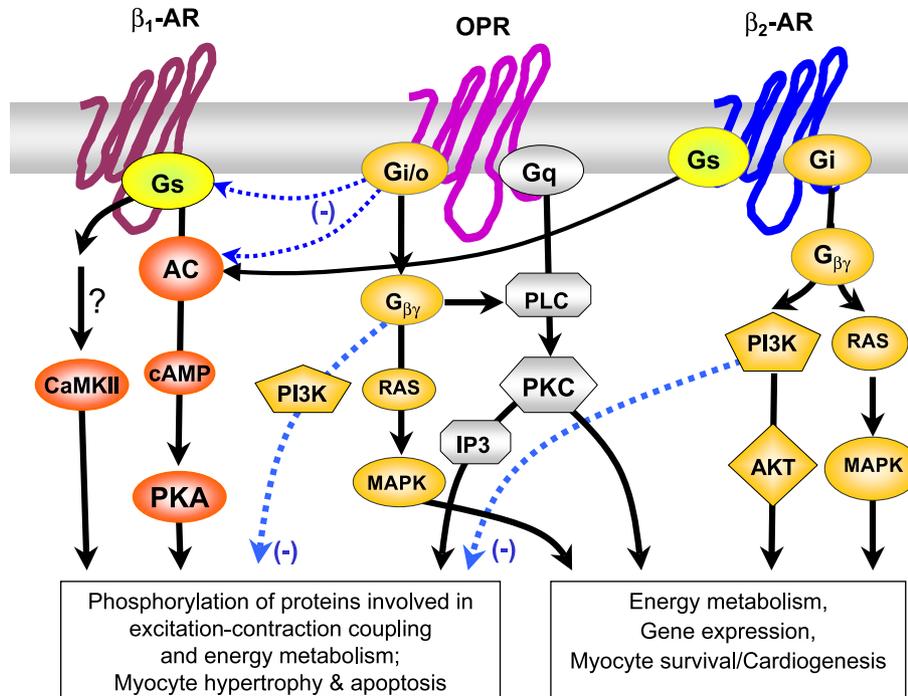


Fig. 1. A simplified scheme of regulatory cross-talk between  $\beta$ -adrenergic receptor ( $\beta$ -AR) subtypes and opioid peptide receptors (OPR) that includes multiple parallel and converging downstream signal transduction pathways. The incidence of receptor heterodimerization is not indicated in the scheme for diagrammatic simplicity. AC: adenylate cyclase; PKA: protein kinase A; PKC: protein kinase C; MAPK: mitogen activated protein kinases; CaMKII:  $\text{Ca}^{2+}$ /calmodulin-dependent protein kinase II;  $G_s$  and  $G_i$  proteins: guanine nucleotide binding proteins with a stimulatory ( $G_s$ ) and inhibitory ( $G_{i/o}$ ) effect on AC, respectively; Gq: G protein q;  $G_{\beta\gamma}$ :  $\beta\gamma$  subunits of G proteins;  $\text{IP}_3$ : inositol 1,4,5-trisphosphate.

archical mechanisms, particularly their distinct G protein coupling and subcellular localization, may render the subtype-specific  $\beta$ -AR/ $\delta$ -OPR interaction.

Regarding the possible molecular and cellular mechanisms responsible for the interaction between OPRs and  $\beta$ -ARs, it has been established that OPR stimulation inhibits  $G_s$  and adenylate cyclase by activating  $G_{i/o}$  signalling pathways, subsequently decreasing cAMP production and PKA activation, leading to a reduction in L-type  $\text{Ca}^{2+}$  current, depletion of  $\text{Ca}^{2+}$  from intracellular pools, and reduced contractility in rat and canine hearts [40–45,65–67]. In addition, OPRs are able to directly regulate voltage-gated  $\text{K}^+$  channel opening and  $\text{Ca}^{2+}$  channel closing without the involvement of the second messenger signalling [68–70]. Moreover, the physical interaction between  $\beta$ -AR and OPR might, at least in part, contribute to the inhibitory effect of OPR stimulation on  $\beta$ -AR-mediated positive inotropic and chronotropic effects in the heart.

## 6. Changes in OPR and $\beta$ -AR signalling in aging heart

Most but not all studies have provided evidence that opioid peptide gene expression and the abundance of opioid peptides are increased with aging in rat hearts [71–73]. However, it is presently unclear whether the functional effects of OPR stimulation in the heart differ with aging.

In contrast, studies over the last three decades have demonstrated that the cardiac response to  $\beta$ -AR stimulation decreases in aging heart [74–77]. Mechanistic studies have been focused on the classic “receptor- $G_s$ -adenylate cyclase-cAMP-PKA” signalling cascade and found multiple defects in the signalling pathway (for review, see Ref. [74]). The suppression of cardiac response to  $\beta$ -AR stimulation is associated with a significant down-regulation of  $\beta$ -AR density and a decrease in the agonist-stimulated adenylate cyclase activity. The age-associated up-regulation of OPR signalling might be, in part, responsible for the reduction in  $\beta$ -AR signalling in aging heart, due to the robust antagonistic effects of stimulation of  $\delta$ -OPR [49,50] or  $\kappa$ -OPR [41–45] on  $\beta$ -AR-mediated positive contractile response.

## 7. $\beta$ -AR and OPR signalling in heart failure

Heart failure induced by a variety of causes is associated with elevated levels of circulating catecholamines, plus a concurrent reduction in  $\beta$ -AR density and desensitization of remaining receptors, leading to a markedly blunted  $\beta$ AR contractile response. Specifically, cardiac contractile response to both  $\beta_1$ - and  $\beta_2$ -AR stimulation is markedly diminished in the failing heart. The reduced  $\beta$ -AR inotropic effect is often accompanied by increased amount or activity of  $G_i$  proteins [78,79] as well as GPCR kinases (GRKs) [80]

and a selective down-regulation of  $\beta_1$ -AR [81]. Recent studies support the notion that  $\beta_1$ - and  $\beta_2$ -AR activate qualitatively and quantitatively different signalling pathways and may play opposing functional roles in the pathogenesis of heart failure. In particular, sustained  $\beta_1$ AR stimulation not only activates the classic cAMP/PKA signalling pathway but also evokes PKA-independent activation of CaMKII [82] promoting cardiac hypertrophy [83,84] and myocyte apoptosis [82,85–88] and selective  $\beta_1$ AR blockers exhibit beneficial effects in patients with CHF [89], whilst enhanced  $\beta_2$ AR activation appears to be cardiac protective [85,90,91] (see Fig. 1). Thus, it is reasonable to speculate that, in the context of heart failure, the selective down-regulation of  $\beta_1$ AR might represent a complementary cardioprotective mechanism to protect myocytes against apoptosis and slow the progression of cardiomyopathy and contractile dysfunction, whereas the up-regulation of  $\beta_2$ AR signalling could be beneficial due to its contractile support and anti-apoptotic effects. These insights also reveal a potential cell logic for the differential interaction of  $\delta$ -OPR with  $\beta$ -AR subtypes:  $\beta_1$ -AR, but not  $\beta_2$ -AR, signalling is negated by  $\delta$ -OPR activation in rat heart [49] (see also Fig. 1).

As discussed earlier, enkephalins negate sympathetic actions on the heart [49–55], in addition to their direct neurally independent negative inotropic effect [40–45]. This might represent a cardioprotective mechanism in the early stage of heart failure to diminish cardiac responsiveness to sympathetic stimulation, thus reducing oxygen demand by limiting work performance. However, exaggerated OPR signalling could contribute to the phenotype of heart failure. For instance, increased levels of Met-enkephalin are correlated with the degree of severity of the disease [92–94]. Further, naloxone is able to improve systemic hemodynamics and myocardial contractile function in a pacing-induced canine heart failure model [95]. Similarly, the heightened opioid peptide activity limits sympathetic activation, whilst inhibition of OPR improves cardiac performance in canine congestive failing hearts [96–97]. The stimulatory effects of the OPR antagonist, naloxone, on the heart could be mediated by an action within the central nervous system [98] as well as a direct inotropic effect [99]. Thus, endogenous enkephalins appear to play an important role in mediating the myocardial depression that occurs in heart failure, and that there may be a role for targeting enkephalins in the clinical therapeutic management of heart failure.

## 8. Ischemic preconditioning and OPR stimulation

Ischemic preconditioning (IPC) is the phenomenon whereby brief sublethal periods of ischemia protect the heart against a more sustained ischemic event. IPC can be divided into two phases: early preconditioning, occurring immediately after the initial stimulus but with a limited duration of 1–2 h, and late preconditioning, occurring

approximately 24 h after the initial stimulus with a duration of almost 72 h [100]. IPC has been shown in both humans [101] and animals [102,103] to reduce infarct size and improve functional recovery after a prolonged period of ischemia.

Considerable evidence points to cardiac mitochondria as contributing an important role in cardioprotection against ischemia–reperfusion injury. The myocardium has a high metabolic rate and is therefore highly dependent on mitochondria for ATP production [104]. Conditions associated with ischemia, prolonged hypoxia and/or deprivation of substrates can ultimately lead to mitochondria-dependent cell death. A key mechanism of action in IPC involves ATP-sensitive potassium channels ( $K_{ATP}^+$  channels). Overexpression of recombinant  $K_{ATP}^+$  channel subunits or pharmacological stimulation of the channels has been shown to promote cytoprotection [104]. Although initially it was suggested that cell surface  $K_{ATP}^+$  channels were responsible for this effect, increasing evidence suggests that *mitochondrial*  $K_{ATP}^+$  channels, which are pharmacologically and histochemically distinct from sarcolemmal  $K_{ATP}^+$  channels [105–109], are the key channels in this process. Opening of mitochondrial  $K_{ATP}^+$  channels after protein kinase C activation and translocation has been shown to be crucial for cardioprotection [110,111].

Multiple lines of evidence suggest that blocking OPR with naloxone or naltrindole can abolish the effects of IPC in humans [112] and rats [113], indicating that  $\delta$ -OPR activation is involved in IPC. This suggests that  $\delta$ -OPR stimulation by endogenous enkephalins may play a major role in IPC [114–118] and may have significant impact on cardiac protection and organ preservation for transplantation in the clinical arena. Recent studies have shown that, similar to  $\delta$ -OPR signalling,  $\kappa$ -OPR stimulation plays an important role in IPC-induced cardiac protection. In fact,  $\kappa$ -OPR is involved in IPC-induced ameliorating effects on arrhythmia as well as infarct, whereas  $\delta$ -OPR activation is only involved in IPC-mediated anti-arrhythmia in perfused rat hearts [119]. The protective effects of  $\kappa$ -OPR stimulation are dependent on activation of PKC and  $K_{ATP}^+$  channels [119].

The cardioprotective signalling pathways linking OPR signalling to mitochondrial  $K_{ATP}^+$  channel activation, and other subcellular targets as seen after PKC isomer activation, awaits future investigation. However, a notable recent breakthrough is the demonstration that numerous types of G-protein-coupled receptors (including opioid receptors), when activated elicit cell protection by signalling via PKB/Akt, mTOR/p70s6k, PI3K, PKC, or PKA pathways which converge to ultimately inhibit GSK-3 $\beta$ . The inhibition of GSK-3 $\beta$  impacts the mitochondrial permeability transition pore complex, to limit induction of mitochondrial pore opening and permit survival following cellular stress [120,121]. As strict metabolic regulation and repair/growth mechanisms are crucial requirements for survival, it is likely

that these pathways are also converged or shared, at least in part, with those involved with the adaptive processes active in aged, hypertrophic or failing hearts.

## 9. Involvement of OPR stimulation in cardiogenesis

Emerging evidence indicates that the prodynorphin gene and its product, dynorphin B, play an important role in cardiogenesis. It has been shown that P19 embryonal pluripotent stem cells are able to express the prodynorphin gene and produce dynorphin B, and that stimulation of  $\kappa$ -OPRs with dynorphin enables these cells to express GATA-4 and Nkx-2.5 genes, key transcription factor-encoding genes essential for cardiogenesis and expression of  $\alpha$ -myosin heavy chain and myosin light chain-2 V genes, two markers of cardiac differentiation [122]. More recently, studies from the same laboratory have further elucidated that in GTR1 embryonic stem cells, stimulation of  $\kappa$ -OPRs promotes cardiogenesis which is associated with activation of PKC- $\delta$  and - $\epsilon$  mainly at the nuclear level and translocation of PKC- $\alpha$ , - $\beta_1$ , and - $\beta_2$  isozymes from cytosol to nucleus, suggesting PKC signalling constitutes the major molecular and cellular mechanism underlying OPR-mediated cardiogenesis in GTR1 embryonic stem cells [123,124]. This notion is further supported by the fact that inhibition of OPR by receptor antagonist reduces cardiomyocyte yield and that PKC inhibitors block the expression of cardiogenic genes and dynorphin B production in the embryonic stem cells and prevent their in vitro differentiation into beating cardiomyocytes [123,124]. Taken together, these recent studies indicate that OPR stimulation regulates cardiogenesis via an autocrine loop signalling mechanism sequentially involving  $\kappa$ -OPR activation by the endogenous agonist, dynorphin B, an increase in PKC activity in the nucleus and expression of cardiac progenitor genes.

In addition, emerging evidence suggests that OPR stimulation serves as a negative growth regulator in renewing and regenerating epithelia, and that disrupting OPR signalling promotes basal cell proliferation [125]. Furthermore,  $\delta$ -OPR stimulation can abolish  $\beta_2$ -AR-mediated cell proliferative effects, indicating that a cross-talk occurs between  $\delta$ -OR and  $\beta_2$ -AR signalling in regulating cell proliferation rate [126].

## 10. Conclusion

Opioid peptides, in particular, enkephalins, have been investigated extensively in the central nervous system. Recently, a novel role of opioids as modulators of cardiac function has emerged. Stimulation of  $\delta$ - and  $\kappa$ -OPR not only exhibits a cardioprotective effect against ischemic and hypoxic injury and directly suppresses cardiac excitation–contraction coupling, but also markedly negates  $\beta$ -AR-mediated positive inotropic effects. The cross-talk between

the two functionally opposite GPCR families might have important physiological and pathological relevance in cardiac aging, heart failure, and cardiac responses to ischemic stress. The exact mechanisms underlying the interaction of these two GPCR systems merit further investigation.

## References

- [1] Rubin GM, Yandell MD, Wortman JR, et al. Comparative genomics of the eukaryotes. *Science* 2000;287:2204–15.
- [2] Kosterlitz HW, Hughes J. Some thoughts on the significance of enkephalin, the endogenous ligand. *Life Sci* 1975;17:91–6.
- [3] Hughes J, Smith TW, Kosterlitz HW, Fothergill LA, Morgan BA, Morris HR. Identification of two related pentapeptides from the brain with potent opiate agonist activity. *Nature* 1975;258:577–80.
- [4] Holaday JW. Cardiovascular effects of endogenous opiate systems. *Annu Rev Pharmacol Toxicol* 1983;23:541–94.
- [5] Tang J, Yang HY, Costa E. Distribution of met5-enkephalin-Arg6-Phe7 (MEAP) in various tissues of rats and guinea pigs. *Life Sci* 1982;31:2303–6.
- [6] Udenfriend S, Kilpatrick DL. Biochemistry of the enkephalins and enkephalin-containing peptides. *Arch Biochem Biophys* 1983;221:309–23.
- [7] Barron BA. Opioid peptides and the heart. *Cardiovasc Res* 1999;43:13–6.
- [8] Jackson KE, Farias M, Stanfill A, Caffrey JL. Delta opioid receptors inhibit vagal bradycardia in the sinoatrial node. *J Cardiovasc Pharmacol Ther* 2001;6:385–93.
- [9] Wittert G, Hope P, Pyle D. Tissue distribution of opioid receptor gene expression in the rat. *Biochem Biophys Res Commun* 1996;218:877–81.
- [10] Ventura C, Capogrossi M, Lakatta E. Myocardial function and endogenous opioids. In: Negri M, Lotti G, Grossman A, editors. *Clinical Perspectives in Endogenous Opioid Production*. Wiley, Chichester, New York; 1992. p. 393–406.
- [11] Giles TD, Sander GE. Mechanism of the cardiovascular response to systemic intravenous administration of leucine-enkephalin in the conscious dog. *Peptides* 1983;4:171–5.
- [12] Feuerstein G, Siren AL. The opioid system in cardiac and vascular regulation of normal and hypertensive states. *Circulation* 1987;75:1125–9.
- [13] Ventura C, Pintus G, Tadolini B. Opioid peptide gene expression in the myocardial cell. *Trends Cardiovasc Med* 1998;8:102–10.
- [14] Wilson SP, Klein RL, Chang KJ, Gasparis MS, Viveros OH, Yang WH. Are opioid peptides co-transmitters in noradrenergic vesicles of sympathetic nerves? *Nature* 1980;288:707–9.
- [15] Springhorn JP, Claycomb WC. Translation of heart preproenkephalin mRNA and secretion of enkephalin peptides from cultured cardiac myocytes. *Am J Physiol* 1992;263:H1560–6.
- [16] Weil J, Eschenhagen T, Fleige G, Mittmann C, Orthey E, Scholz H. Localization of preproenkephalin mRNA in rat heart: selective gene expression in left ventricular myocardium. *Am J Physiol* 1998;275:H378–84.
- [17] Howells RD, Kilpatrick DL, Bailey LC, Noe M, Udenfriend S. Proenkephalin mRNA in rat heart. *Proc Natl Acad Sci U S A* 1986;83:1960–3.
- [18] Low KG, Allen RG, Melner MH. Association of proenkephalin transcripts with polyribosomes in the heart. *Mol Endocrinol* 1990;4:1408–14015.
- [19] Younes A, Pepe S, Barron BA, Spurgeon HA, Lakatta EG, Caffrey JL. Cardiac synthesis, processing, and coronary release of enkephalin-related peptides. *Am J Physiol Heart Circ Physiol* 2000;279:H1989–98.

- [20] Semmoum Y, Younes A, Coudert J. Effects of ischemia on the metabolism of cardiac enkephalins. *Arch Physiol Biochem* 2001;109:18–23.
- [21] Ventura C, Guarnieri C, Vaona I, Campana G, Pintus G, Spampinato S. Dynorphin gene expression and release in the myocardial cell. *J Biol Chem* 1994;269:5384–6.
- [22] Chavkin C, James IF, Goldstein A. Dynorphin is a specific endogenous ligand of the  $\kappa$  opioid receptor. *Science* 1982;215:413–5.
- [23] Ventura C, Pintus G, Vaona I, Bennardini F, Pinna G, Tadolini B. Phorbol ester regulation of opioid peptide gene expression in myocardial cells. Role of nuclear protein kinase. *J Biol Chem* 1995;270:30115–20.
- [24] Ventura C, Pintus G, Fiori MG, Bennardini F, Pinna G, Gaspa L. Opioid peptide gene expression in the primary hereditary cardiomyopathy of the Syrian hamster: I. Regulation of prodynorphin gene expression by nuclear protein kinase C. *J Biol Chem* 1997;272:6685–92.
- [25] Ventura C, Pintus G. Opioid peptide gene expression in the primary hereditary cardiomyopathy of the Syrian hamster: III. Autocrine stimulation of prodynorphin gene expression by dynorphin B. *J Biol Chem* 1997;272:6699–705.
- [26] Dhawan BN, Cesselin F, Raghbir R, Reisine T, Bradley PB, Portoghesi PS, et al. International Union of Pharmacology: XII. Classification of opioid receptors. *Pharmacol Rev* 1996;48:567–92.
- [27] Chen Y, Mestek A, Liu J, Hurley JA, Yu L. Molecular cloning and functional expression of a  $\mu$ -opioid receptor from rat brain. *Mol Pharmacol* 1993;44:8–12.
- [28] Meng F, Xie GX, Thompson RC, Mansour A, Goldstein A, Watson SJ, et al. Cloning and pharmacological characterization of a rat  $\kappa$  opioid receptor. *Proc Natl Acad Sci U S A* 1993;90:9954–8.
- [29] Kieffer BL, Befort K, Gaveriaux-Ruff C, Hirth CG. The  $\delta$ -opioid receptor: isolation of a cDNA by expression cloning and pharmacological characterization. *Proc Natl Acad Sci U S A* 1992;89:12048–52.
- [30] Evans CJ, Keith Jr DE, Morrison H, Magendzo K, Edwards RH. Cloning of a  $\delta$  opioid receptor by functional expression. *Science* 1992;258:1952–5.
- [31] Reisine T, Bell GI. Molecular biology of opioid receptors. *Trends Neurosci* 1993;16:506–10.
- [32] Yasuda K, Raynor K, Kong H, Breder CD, Takeda J, Reisne T, et al. Cloning and functional comparison of  $\kappa$  and  $\delta$  opioid receptors from mouse brain. *Neurobiology* 1993;90:6736–40.
- [33] Krumins SA, Faden AI, Feuerstein G. Opiate binding in rat hearts: modulation of binding after hemorrhagic shock. *Biochem Biophys Res Commun* 1985;127:120–8.
- [34] Ventura C, Bastagli L, Bernardi P, Calderera CM, Guarnieri C. Opioid receptors in rat cardiac sarcolemma: effect of phenylephrine and isoproterenol. *Biochim Biophys Acta* 1989;987:69–74.
- [35] Tai KK, Jin WQ, Chan TK, Wong TM. Characterization of [3H]U69593 binding sites in the rat heart by receptor binding assays. *J Mol Cell Cardiol* 1991;23:1297–302.
- [36] Zhang WM, Jin WQ, Wong TM. Multiplicity of kappa opioid receptor binding in the rat cardiac sarcolemma. *J Mol Cell Cardiol* 1996;28:1547–54.
- [37] Zimlichman R, Gefel D, Eliahou H, Matas Z, Rosen B, Gass S, et al. Expression of opioid receptors during heart ontogeny in normotensive and hypertensive rats. *Circulation* 1996;93:1020–5.
- [38] Valtchanova-Matchouganska A, Ojewole JA. Involvement of opioid  $\delta$ - and  $\kappa$ -receptors in ischemic preconditioning in a rat model of myocardial infarction. *Methods Find Exp Clin Pharmacol* 2002;24:139–44.
- [39] Sofuoglu M, Portoghesi PS, Takemore AE. Differential antagonism of delta opioid agonists by naltrindole and its benzofuran analog (NTB) in mice: evidence for delta opioid receptor subtypes. *J Pharmacol Exp Ther* 1991;257:767–80.
- [40] Xiao RP, Spurgeon HA, Capogrossi MC, Lakatta EG. Stimulation of opioid peptide receptors on cardiac myocytes reduces L-type Ca channel current. *J Mol Cell Cardiol* 1993;25:661–6.
- [41] Ventura C, Spurgeon H, Lakatta EG, Guarnieri C, Capogrossi MC.  $\kappa$  and  $\delta$  opioid receptor stimulation affects cardiac myocyte function and  $Ca^{2+}$  release from an intracellular pool in myocytes and neurons. *Circ Res* 1992;70:66–81.
- [42] Ventura C, Capogrossi MC, Spurgeon HA, Lakatta EG.  $\kappa$ -Opioid peptide receptor stimulation increases cytosolic pH and myofilament responsiveness to  $Ca^{2+}$  in cardiac myocytes. *Am J Physiol* 1991;261:H1671–4.
- [43] Bian JS, Wang HX, Zhang WM, Wong TM. Effects of  $\kappa$ -opioid receptor stimulation in the heart and the involvement of protein kinase C. *Br J Pharmacol* 1998;124:600–6.
- [44] Sheng JZ, Wong NS, Tai KK, Wong TM. Lithium attenuates the effects of dynorphin A(1–13) on inositol 1,4,5-trisphosphate and intracellular  $Ca^{2+}$  in rat ventricular myocytes. *Life Sci* 1996;59:2181–6.
- [45] Sheng JZ, Wong NS, Wang HX, Wong TM. Pertussis toxin, but not tyrosine kinase inhibitors, abolishes effects of U-50488H on  $[Ca^{2+}]_i$  in myocytes. *Am J Physiol* 1997;272:C560–4.
- [46] Jordan BA, Devi LA. G-protein-coupled receptor heterodimerization modulates receptor function. *Nature* 1999;399:697–700.
- [47] Portoghesi PS, Lunzer MM. Identity of the putative  $\delta_1$ -opioid receptor as a  $\delta$ - $\kappa$  heteromer in the mouse spinal cord. *Eur J Pharmacol* 2003;467:233–4.
- [48] Jordan BA, Trapaidze N, Gomes I, Nivarthi R, Devi LA. Oligomerization of opioid receptors with  $\beta_2$ -adrenergic receptors: a role in trafficking and mitogen-activated protein kinase activation. *Proc Natl Acad Sci U S A* 2001;98:343–8.
- [49] Pepe S, Xiao RP, Hohl C, Altschuld R, Lakatta EG. ‘Cross talk’ between opioid peptide and adrenergic receptor signaling in isolated rat heart. *Circulation* 1997;95:2122–9.
- [50] Xiao RP, Pepe S, Spurgeon HA, Capogrossi MC, Lakatta EG. Opioid peptide receptor stimulation reverses  $\beta$ -adrenergic effects in rat heart cells. *Am J Physiol* 1997;272:H797–805.
- [51] Yu XC, Li HY, Wang HX, Wong TM. U50,488H inhibits effects of norepinephrine in rat cardiomyocytes—cross-talk between  $\kappa$ -opioid and  $\beta$ -adrenergic receptors. *J Mol Cell Cardiol* 1998;30:405–13.
- [52] Yu XC, Wang HX, Pei JM, Wong TM. Anti-arrhythmic effect of  $\kappa$ -opioid receptor stimulation in the perfused rat heart: involvement of a cAMP-dependent pathway. *J Mol Cell Cardiol* 1999;31:1809–19.
- [53] Shan J, Yu XC, Fung ML, Wong TM. Attenuated ‘‘cross talk’’ between  $\kappa$ -opioid receptors and  $\beta$ -adrenoceptors in the heart of chronically hypoxic rats. *Pflügers Arch* 2002;444:126–32.
- [54] Yu XC, Wang HX, Zhang WM, Wong TM. Cross-talk between cardiac  $\kappa$ -opioid and  $\beta$ -adrenergic receptors in developing hypertensive rats. *J Mol Cell Cardiol* 1999;31:597–605.
- [55] Wong TM, Shan J. Modulation of sympathetic actions on the heart by opioid receptor stimulation. *J Biomed Sci* 2001;8:299–306.
- [56] Aprigliano O, Rybin VO, Pak E, Robinson RB, Steinberg SF.  $\beta_1$ - and  $\beta_2$ -Adrenergic receptors exhibit differing susceptibility to muscarinic accentuated antagonism. *Am J Physiol* 1997;272:H2726–35.
- [57] Rybin VO, Xu X, Lisanti MP, Steinberg SF. Differential targeting of  $\beta$ -adrenergic receptor subtypes and adenylyl cyclase to cardiomyocyte caveolae. A mechanism to functionally regulate the cAMP signaling pathway. *J Biol Chem* 2000;275:41447–57.
- [58] Feron O, Smith TW, Michel T, Kelly RA. Dynamic targeting of the agonist-stimulated  $m_2$  muscarinic acetylcholine receptor to caveolae in cardiac myocytes. *J Biol Chem* 1997;272:17744–8.
- [59] Xiao RP, Ji X, Lakatta EG. Functional coupling of the  $\beta_2$ -adrenoceptor to a pertussis toxin-sensitive G protein in cardiac myocytes. *Mol Pharmacol* 1995;47:322–9.
- [60] Xiao RP, Avdonin P, Zhou YY, Cheng H, Akhter SA, Eschenhagen T, et al. Coupling of  $\beta_2$ -adrenoceptor to  $G_i$  proteins and its physiological relevance in murine cardiac myocytes. *Circ Res* 1999;84:43–52.
- [61] Kiltz JD, Gerhardt MA, Richardson MD, Sreeram G, Mackensen

- GB, Grocott HP, et al.  $\beta_2$ -Adrenergic and several other G protein-coupled receptors in human atrial membranes activate both  $G_s$  and  $G_i$ . *Circ Res* 2000;87:705–9.
- [62] Kuschel M, Zhou YY, Cheng H, Zhang SJ, Chen-Izu Y, Lakatta EG, et al.  $G_i$  protein-mediated functional compartmentalization of cardiac  $\beta_2$ -adrenergic signaling. *J Biol Chem* 1999;274:22048–52.
- [63] Gong H, Sun H, Koch WJ, Rau T, Eschenhagen T, Ravens U, et al. Specific  $\beta_2$ AR blocker ICI 118,551 actively decreases contraction through a  $G_i$ -coupled form of the  $\beta_2$ AR in myocytes from failing human heart. *Circulation* 2002;105:2497–503.
- [64] Cerbai E, Pino R, Rodriguez ML, Mugelli A. Modulation of the pacemaker current if by  $\beta$ -adrenoceptor subtypes in ventricular myocytes isolated from hypertensive and normotensive rats. *Cardiovasc Res* 1999;42:121–9.
- [65] Yu XC, Diao TM, Pei JM, Zhang WM, Wong NS, Wong TM.  $\kappa$ -Opioid receptor agonist inhibits the cholera toxin-sensitive G protein in the heart. *J Cardiovasc Pharmacol* 2001;38:232–9.
- [66] Niroomand F, Mura RA, Piacentini L, Kubler W. Opioid receptor agonists activate pertussis toxin-sensitive G proteins and inhibit adenylyl cyclase in canine cardiac sarcolemma. *Naunyn-Schmiedeberg Arch Pharmacol* 1996;354:643–9.
- [67] Schultz JE, Gross GJ. Opioids and cardioprotection. *Pharmacol Ther* 2001;89:123–37.
- [68] Wild KD, Vanderah T, Mosberg HI, Porreca F. Opioid  $\delta$  receptor subtypes are associated with different potassium channels. *Eur J Pharmacol* 1991;193:135–6.
- [69] Gross RA, Moises HC, Uhler MD, Macdonald RL. Dynorphin A and cAMP-dependent protein kinase independently regulate neuronal calcium currents. *Proc Natl Acad Sci U S A* 1990;87:7025–9.
- [70] Ulens C, Daenens P, Tytgat J. The dual modulation of GIRK1/GIRK2 channels by opioid receptor ligands. *Eur J Pharmacol* 1999;385:239–45.
- [71] Boluyt MO, Younes A, Caffrey JL, O'Neill L, Barron BA, Crow MT, et al. Age-associated increase in rat cardiac opioid production. *Am J Physiol* 1993;265:H212–8.
- [72] Caffrey JL, Boluyt MO, Younes A, Barron BA, O'Neill L, Crow MT, et al. Aging, cardiac proenkephalin mRNA and enkephalin peptides in the Fisher 344 rat. *J Mol Cell Cardiol* 1994;26:701–11.
- [73] Bhargava HN, Matwyshyn GA, Hanissian S, Tejwani GA. Opioid peptides in pituitary gland, brain regions and peripheral tissues of spontaneously hypertensive and Wistar–Kyoto normotensive rats. *Brain Res* 1988;440:333–40.
- [74] Lakatta EG. Arterial and cardiac aging: major shareholders in cardiovascular disease enterprises: Part III. Cellular and molecular clues to heart and arterial aging. *Circulation* 2003;107:490–7.
- [75] Xiao RP, Spurgeon HA, O'Connor F, Lakatta EG. Age-associated changes in  $\beta$ -adrenergic modulation on rat cardiac excitation–contraction coupling. *J Clin Invest* 1994;94:2051–9.
- [76] Xiao RP, Tomhave ED, Ji X, Boluyt MO, Cheng H, Lakatta EG, et al. Age-associated reductions in cardiac  $\beta_1$ - and  $\beta_2$ -adrenoceptor responses without changes in inhibitory G proteins or receptor kinases. *J Clin Invest* 1998;101:1273–82.
- [77] Cerbai E, Guerra L, Varani K, Barbieri M, Borea PA, Mugelli A.  $\beta$ -Adrenoceptor subtypes in young and old rat ventricular myocytes: a combined patch-clamp and binding study. *Br J Pharmacol* 1995;116:1835–42.
- [78] Eschenhagen T, Mende U, Nose M, Schmitz W, Scholz H, Haverich A, et al. Increased messenger RNA level of the inhibitory G protein  $\alpha$  subunit  $G_{i\alpha 1}$  in human end-stage heart failure. *Circ Res* 1992;70:688–96.
- [79] Bohm M, Eschenhagen T, Gierschik P, Larisch K, Lensche H, Mende U, et al. Radioimmunochemical quantification of  $G_{i\alpha}$  in right and left ventricles from patients with ischaemic and dilated cardiomyopathy and predominant left ventricular failure. *J Mol Cell Cardiol* 1994;26:133–49.
- [80] Hammond HK. Mechanisms for myocardial  $\beta$ -adrenergic receptor desensitization in heart failure. *Circulation* 1993;87:652–4.
- [81] Bristow MR, Ginsburg R, Umans V, Fowler M, Minobe W, Rasmussen R, et al.  $\beta_1$ - and  $\beta_2$ -Adrenergic-receptor subpopulations in nonfailing and failing human ventricular myocardium: coupling of both receptor subtypes to muscle contraction and selective  $\beta_1$ -receptor down-regulation in heart failure. *Circ Res* 1986;59:297–309.
- [82] Zhu WZ, Wang SQ, Chakir K, Kolbilka BK, Cheng H, Xiao RP. Linkage of  $\beta_1$ -adrenergic stimulation to apoptotic heart cell death through protein kinase A-independent activation of  $Ca^{2+}$ /calmodulin kinase II. *J Clin Invest* 2003;111:617–25.
- [83] Morisco C, Zebrowski DC, Vatner DE, Vatner SF, Sadoshima J.  $\beta$ -Adrenergic cardiac hypertrophy is mediated primarily by the  $\beta_1$ -subtype in the rat heart. *J Mol Cell Cardiol* 2001;33:561–73.
- [84] Schafer M, Frischkopf K, Taimor G, Piper HM, Schluter KD. Hypertrophic effect of selective  $\beta_1$ -adrenoceptor stimulation on ventricular cardiomyocytes from adult rat. *Am J Physiol Cell Physiol* 2000;279:C495–503.
- [85] Communal C, Singh K, Sawyer DB, Colucci WS. Opposing effects of  $\beta_1$ - and  $\beta_2$ -adrenergic receptors on cardiac myocyte apoptosis: role of a pertussis toxin-sensitive G protein. *Circulation* 1999;100:2210–2.
- [86] Zaugg M, Xu W, Lucchinetti E, Shafiq SA, Jamali NZ, Siddiqui MAQ.  $\beta$ -Adrenergic receptor subtypes differentially affect apoptosis in adult rat ventricular myocytes. *Circulation* 2000;102:344–50.
- [87] Engelhardt S, Hein L, Wiesmann F, Lohse MJ. Progressive hypertrophy and heart failure in  $\beta_1$ -adrenergic receptor transgenic mice. *Proc Natl Acad Sci U S A* 1999;96:7059–64.
- [88] Metra M, Giubbini R, Nodari S, Boldi E, Modena MG, Dei Cas L. Differential effects of  $\beta$ -blockers in patients with heart failure: a prospective, randomized, double-blind comparison of the long-term effects of metoprolol versus carvedilol. *Circulation* 2000;102:546–51.
- [89] Bristow MR. Mechanistic and clinical rationales for using  $\beta$ -blockers in heart failure. *J Card Fail* 2000;6:8–14.
- [90] Chesley A, Lundberg MS, Asai T, Xiao RP, Ohtani S, Lakatta EG, et al.  $\beta_2$ -Adrenergic receptor delivers an anti-apoptotic signal to cardiac myocytes through  $G_i$ -dependent signaling pathways. *Circ Res* 2000;87:1172–9.
- [91] Zhu WZ, Zheng M, Koch WJ, Lefkowitz RJ, Kobilka BK, Xiao RP. Dual modulation of cardiac cell survival and cell death by  $\beta_2$ -adrenergic signaling in adult mouse heart cells. *Proc Natl Acad Sci U S A* 2001;98:1607–12.
- [92] Lowe H. Role of endogenous opioids in heart failure. *Z Kardiol* 1991;80:47–51.
- [93] Fontana F, Bernardi P, Pich EM, Capelli M, Bortoluzzi L, Spampinato S, et al. Relationship between plasma atrial natriuretic factor and opioid peptide levels in healthy subjects and in patients with acute congestive heart failure. *Eur Heart J* 1993;14:219–25.
- [94] Imai N, Kashiki M, Woolf PD, Liang CS. Comparison of cardiovascular effects of  $\mu$ - and  $\delta$ -opioid receptor antagonists in dogs with congestive heart failure. *Am J Physiol* 1994;267:H912–7.
- [95] Himura Y, Liang CS, Imai N, Delehanty JM, Woolf PD, Hood Jr WB. Short-term effects of naloxone on hemodynamics and baroreflex function in conscious dogs with pacing-induced congestive heart failure. *J Am Coll Cardiol* 1994;23:194–200.
- [96] Liang CS, Imai N, Stone CK, Woolf PD, Kawashima S, Tuttle RR. The role of endogenous opioids in congestive heart failure: effects of nalmefene on systemic and regional hemodynamics in dogs. *Circulation* 1987;75:443–51.
- [97] Yatani A, Imai N, Himura Y, Suematsu M, Liang CS. Chronic opiate-receptor inhibition in experimental congestive heart failure in dogs. *Am J Physiol* 1997;272:H478–84.
- [98] Sakamoto S, Stone CK, Woolf PD, Liang CS. Opiate receptor antagonism in right-sided congestive heart failure. Naloxone exerts salutary hemodynamic effects through its action on the central nervous system. *Circ Res* 1989;65:103–14.
- [99] Llobel F, Laorden ML. Effects of  $\mu$ -,  $\delta$ - and  $\kappa$ -opioid antagonists in

- atrial preparations from nonfailing and failing human hearts. *Gen Pharmacol* 1997;28:371–4.
- [100] Sommerschild HT, Kirkeboen KA. Preconditioning—endogenous defence mechanisms of the heart. *Acta Anaesthesiol Scand* 2002; 46:123–37.
- [101] Abete P, Ferrara N, Cacciatore F, Madrid A, Bianco S, Calabrese C, et al. Angina-induced protection against myocardial infarction in adult and elderly patients: a loss of preconditioning mechanism in the aging heart? *J Am Coll Cardiol* 1997;30:947–54.
- [102] Karck M, Tanaka S, Bolling SF, Simon A, Su TP, Oeltgen PR, et al. Myocardial protection by ischemic preconditioning and delta-opioid receptor activation in the isolated working rat heart. *J Thorac Cardiovasc Surg* 2001;122:986–92.
- [103] Laclau MN, Boudina S, Thambo JB, Tariosse L, Gouverneur G, Bonoron-Adele S, et al. Cardioprotection by ischemic preconditioning preserves mitochondrial function and functional coupling between adenine nucleotide translocase and creatine kinase. *J Mol Cell Cardiol* 2001;33:947–56.
- [104] Ozcan C, Holmuhamedov EL, Jahangir A, Terzic A. Diazoxide protects mitochondria from anoxic injury: implications for myopreservation. *J Thorac Cardiovasc Surg* 2001;121:298–306.
- [105] Akao M, Ohler A, O'Rourke B, Marban E. Mitochondrial ATP-sensitive potassium channels inhibit apoptosis induced by oxidative stress in cardiac cells. *Circ Res* 2001;88:1267–75.
- [106] Hu H, Sato T, Seharaseyon J, Liu Y, Johns DC, O'Rourke B, et al. Pharmacological and histochemical distinctions between molecularly defined sarcolemmal KATP channels and native cardiac mitochondrial KATP channels. *Mol Pharmacol* 1999;55: 1000–5.
- [107] Sasaki N, Murata M, Guo Y, Jo SH, Ohler A, Akao M, et al. MCC-134, a single pharmacophore, opens surface ATP-sensitive potassium channels, blocks mitochondrial ATP-sensitive potassium channels, and suppresses preconditioning. *Circulation* 2003;107: 1183–8.
- [108] Katoh H, Nishigaki N, Hayashi H. Diazoxide opens the mitochondrial permeability transition pore and alters  $Ca^{2+}$  transients in rat ventricular myocytes. *Circulation* 2002;105:2666–71.
- [109] Sato T, Sasaki N, Seharaseyon J, O'Rourke B, Marban E. Selective pharmacological agents implicate mitochondrial but not sarcolemmal KATP channels in ischemic cardioprotection. *Circulation* 2000; 101:2418–23.
- [110] Fryer RM, Wang Y, Hsu AK, Gross GJ. Essential activation of PKC- $\delta$  in opioid-initiated cardioprotection. *Am J Physiol Heart Circ Physiol* 2001;280:H1346–53.
- [111] Sato T, O'Rourke B, Marban E. Modulation of mitochondrial ATP-dependent  $K^+$  channels by protein kinase C. *Circ Res* 1998;83: 110–4.
- [112] Tomai F, Crea F, Gaspardone A, Versaci F, Ghini AS, Ferri C, et al. Effects of naloxone on myocardial ischemic preconditioning in humans. *J Am Coll Cardiol* 1999;33:1863–9.
- [113] Huh J, Gross GJ, Nagase H, Liang BT. Protection of cardiac myocytes via  $\delta_1$ -opioid receptors, protein kinase C, and mitochondrial KATP channels. *Am J Physiol Heart Circ Physiol* 2001;280: H377–83.
- [114] Bell SP, Sack MN, Patel A, Opie LH, Yellon DM.  $\delta$  Opioid receptor stimulation mimics ischemic preconditioning in human heart muscle. *J Am Coll Cardiol* 2000;36:2296–302.
- [115] Takasaki Y, Wolff RA, Chien GL, van Winkle DM. Met5-enkephalin protects isolated adult rabbit cardiomyocytes via delta-opioid receptors. *Am J Physiol* 1999;277:H2442–50.
- [116] Sigg DC, Coles JA, Gallagher WJ, Oeltgen PR, Iaizzo PA. Opioid preconditioning: myocardial function and energy metabolism. *Ann Thorac Surg* 2001;72:1576–82.
- [117] Bolling SF, Badhwar V, Schwartz CF, Oeltgen PR, Kilgore K, Su TP. Opioids confer myocardial tolerance to ischemia: interaction of  $\delta$  opioid agonists and antagonists. *J Thorac Cardiovasc Surg* 2001; 122:476–81.
- [118] Su T. Delta opioid peptide [D-Ala $^2$ , D-Leu $^5$ ]enkephalin promotes cell survival. *J Biomed Sci* 2000;7:195–9.
- [119] Wang GY, Wu S, Pei JM, Yu XC, Wong TM.  $\kappa$ - but not  $\delta$ -opioid receptors mediate effects of ischemic preconditioning on both infarct and arrhythmia in rats. *Am J Physiol Heart Circ Physiol* 2001;280: H384–91.
- [120] Juhaszova M, Zorov D, Kim S, Pepe S, Fu Q, Fishbein K, et al. GSK-3 $\beta$  mediates convergence of protection signaling to inhibit the mitochondrial permeability transition pore. *J Clin Invest* 2004 [in press].
- [121] Gross ER, Hsu AK, Gross GJ. Opioid-induced cardioprotection occurs via glycogen synthase kinase  $\beta$  inhibition during reperfusion in intact rat hearts. *Circ Res* 2004;94:960–6.
- [122] Ventura C, Maioli M. Opioid peptide gene expression primes cardiogenesis in embryonal pluripotent stem cells. *Circ Res* 2000;87: 189–94.
- [123] Ventura C, Zinellu E, Maninchedda E, Maioli M. Dynorphin B is an agonist of nuclear opioid receptors coupling nuclear protein kinase C activation to the transcription of cardiogenic genes in GTR1 embryonic stem cells. *Circ Res* 2003;92:623–9.
- [124] Ventura C, Zinellu E, Maninchedda E, Fadda M, Maioli M. Protein kinase C signaling transduces endorphin-primed cardiogenesis in GTR1 embryonic stem cells. *Circ Res* 2003;92:617–22.
- [125] Wilson RP, McLaughlin PJ, Lang CM, Zagon IS. The opioid growth factor, [Met $^5$ ]-enkephalin, inhibits DNA synthesis during recombination of mouse tail skin. *Cell Prolif* 2000;33:63–73.
- [126] Agarwal D, Glasel JA. Differential effects of opioid and adrenergic agonists on proliferation in a cultured cell line. *Cell Prolif* 1999;32: 215–29.